# Package: adwave (via r-universe)

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Title Wavelet Analysis of Genomic Data from Admixed Populations

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Description Implements wavelet-based approaches for describing population admixture. Principal Components Analysis (PCA) is used to define the population structure and produce a localized admixture signal for each individual. Wavelet summaries of the PCA output describe variation present in the data and can be related to population-level demographic processes. For more details, see J Sanderson, H Sudoyo, TM Karafet, MF Hammer and MP Cox. 2015. Reconstructing past admixture processes from local genomic ancestry using wavelet transformation. Genetics 200:469-481 <doi:10.1534 genetics.115.176842="">.</doi:10.1534>
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admix

Simulated Admixed Population Data

## **Description**

The example dataset provides simulated allele calls for 50 individuals from 3 populations. We consider a simple admixture scenario where two ancestral populations, popA (sample size  $n_A =$  15) and popB (sample size  $n_B =$  15), merged 160 generations ago to form the admixed population, popAB (sample size  $n_{AB} =$  20). The ancestral populations contributed to the admixed population with probability 0.5.

### Usage

data(admix)

#### **Format**

A list with the following entries:

- data: 3,000 x 50 data matrix with 3,000 genotype calls (rows) for 50 individuals (columns). Raw entries take the value 0 if heterozygous, and -1 or 1 if homozygous. Polymorphisms are ordered by their physical positions. Column names list individual IDs, row names list polymorphism IDs.
- map: 3,000 x 2 data matrix providing a mapping to genetic distance ('recombination distance') for each polymorphism.
- colplot: vector of length 50. Plotting color for each individual.
- populations: list of length 3. Character vectors giving the IDs of individuals in each population. IDs must map to the column names of the data matrix.

#### **Details**

Further description of the dataset can be found in Sanderson et al. (2015). The data were simulated using MaCS (Chen et al. 2009).

#### References

Sanderson J, H Sudoyo, TM Karafet, MF Hammer and MP Cox. 2015. Reconstructing past admixture processes from local genomic ancestry using wavelet transformation. *Genetics* 200:469-481. https://doi.org/10.1534/genetics.115.176842

Chen GK, P Marjoram and JD Wall. 2009. Fast and flexible simulation of DNA sequence data. *Genome Research* 19:136-142. https://doi.org/10.1101/gr.083634.108

## **Examples**

```
data(admix)
str(admix)
```

plotsignal 3

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Plot Localized Admixture Signals

# Description

Plotting function for objects of class adsig.

# Usage

```
\label{eq:poph}  \mbox{plotsignal(x, ind = NULL, popA = NULL, popB = NULL, xlab = NULL, ylab = NULL, ylim = NULL, main = NULL) }
```

# Arguments

X	object of class adsig for plotting.
ind	character giving ID of a single individual to plot.
рорА	(optional) name of ancestral population 1.
рорВ	(optional) name of ancestral population 2.
xlab	(optional) character string for x axis label.
ylab	(optional) character string for y axis label.
ylim	(optional) vector giving plotting range for y axis.
main	(optional) character string for title.

## Value

Produces figure.

## Author(s)

Jean Sanderson

## References

Sanderson J, H Sudoyo, TM Karafet, MF Hammer and MP Cox. 2015. Reconstructing past admixture processes from local genomic ancestry using wavelet transformation. *Genetics* 200:469-481. https://doi.org/10.1534/genetics.115.176842

#### See Also

signal

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## **Examples**

```
data(admix)
# Generate the admixture signal with windowing
AdexPCA2 <- signal(admix$data,popA="popA",popB="popB",populations=admix$populations,
    tol=0.001,n.signal=1000,window.size=0.01)
# Plot resulting admixture signal for one individual
plotsignal(AdexPCA2,ind="AD00001",popA=AdexPCA2$popA,popB=AdexPCA2$popB)</pre>
```

signal

Compute Localized Admixture Signals

## **Description**

Produces estimates of localized ancestry for each individual.

## Usage

```
signal(table, who = colnames(table), populations, popA = NA, popB = NA,
normalize = FALSE, n.pca = 5, PCAonly = FALSE, verbose = TRUE, tol = 0.001,
n.signal = NULL, window.size = NULL, genmap = NULL)
```

### **Arguments**

table	matrix of genotype calls (rows, length $T$ ) versus individuals (columns, length $n$ ).
who	individuals to include in the analysis.
populations	list containing a vector of IDs for each population in the analysis.
рорА	name of ancestral population 1 (used for forming the axes of variation). Must match one of the names in populations.
рорВ	name of ancestral population 2 (used for forming the axes of variation). Must match one of the names in populations.
normalize	if TRUE, normalize the data matrix. Default is FALSE.
n.pca	number of PCA axes to compute (only the first principal component is used for forming the signals, but additional components may be desired for visualization). Default is 5.
PCAonly	if TRUE, only compute the PCA, do not compute the signals. Default is FALSE.
verbose	if TRUE, print summary to screen. Default is TRUE.
tol	tolerance for normalization of admixture signals ( $\epsilon$ in accompanying paper). Default is 0.001.
n.signal	(optional) number of data points in the windowed signal.
window.size	(optional) size of window specified as a proportion of total length; e.g., window.size = 0.01 with signal of length $T$ = 3000 SNPs generates windows of $0.01x3000 = 30$ polymorphisms. Value need not be a round number.
genmap	(optional) genetic distance of genotype calls, supplied as vector of length T.

chromosome (rather than physical position).

If specified, signals will be formulated in terms of genetic distance along the

signal 5

#### **Details**

Applies PCA to genome-wide data using ancestral reference populations. The first eigenvector reflects the population structure. All individuals are then projected on to this axis to form the SNP-level admixture signals. PCA scores are used to estimate the proportion of admixture at the level of individuals (indP) and populations (popP). There is no restriction on the length of the data (number of SNPs) and the default is to provide an estimate of localized ancestry at each SNP.

Optionally, it is also possible to window the signals, producing processed signals of length n.signal. The windows may be overlapping or disjoint with width specified through the window.size option (see examples). If genmap is specified, the signals will be formulated in terms of genetic distance along the chromosome (note: this function is not described in the accompanying paper).

#### Value

Returns an object of class adsig, a list with the following components:

call	function call.
date	date of function call.
individuals	individuals for whom projections on the first principal component are calculated.
n.snps	number of polymorphisms in the table.
signals	The admixture signals, output as a $Txn$ data matrix, where $n$ is the number of individuals and $T$ is the number of data points (either the number of polymorphisms if n.signal = NULL or n.signal otherwise).
n.tol	the number of entries replaced by zero in the normalization procedure. This is dependent on the value set for the tolerance, tol.
popP	estimated proportion of admixture for each population.
indP	estimated proportion of admixture for each individual.
pa.ind	columns are principal axes in individual coordinates ( $n_A + n_B$ rows, n.pca columns).
pa.snp	columns are principal axes in polymorphism coordinates (T rows, n.pca columns).
G	matrix of quadratic form in individual coordinates.
ev	vector of eigenvalues.
gendist	(only if genmap is specified in input) Vector of genetic distances along the chromosome, length n.signal.

## Author(s)

Jean Sanderson

## References

Sanderson J, H Sudoyo, TM Karafet, MF Hammer and MP Cox. 2015. Reconstructing past admixture processes from local genomic ancestry using wavelet transformation. *Genetics* 200:469-481. https://doi.org/10.1534/genetics.115.176842

## See Also

wavesum, plotsignal

#### **Examples**

```
data(admix)
# EXAMPLE 1
 # Generate the admixture signal
AdexPCA <- signal(admix$data,popA="popA",popB="popB",populations=admix$populations,tol=0.001,
       n.signal=NULL)
 # Plot the resulting PCA
 plot(AdexPCA$pc.ind[,1],AdexPCA$pc.ind[,2],col=admix$colplot,xlab="PC1",ylab="PC2",pch=16)
 legend("bottomright",c("popA","popB","popAB"),col=c(3,4,2),pch=16)
 # EXAMPLE 2
 # Generate the admixture signal with windowing
AdexPCA2 <- signal(admix$data,popA="popA",popB="popB",populations=admix$populations,tol=0.001,
       n.signal=1000,window.size=0.01)
 # Plot resulting admixture signal for one individual
plotsignal(AdexPCA2,ind="AD00001",popA=AdexPCA2$popA,popB=AdexPCA2$popB)
# EXAMPLE 3
# Generate the admixture signal with windowing
# As in EXAMPLE 2 but with n.signal reduced to 100 to provide disjoint windows
AdexPCA3 <- signal(admix\$data,popA="popA",popB="popB",populations=admix\$populations,tol=0.001,admix\$data,popA="popA",popB="popB",populations=admix\$populations,tol=0.001,admix\$data,popA="popA",popB="popB",populations=admix\$populations,tol=0.001,admix\$data,popA="popA",popB="popB",popB="popB",populations=admix\$populations,tol=0.001,admix\$data,popA="popA",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB="popB="popB="popB",popB="popB="popB="popB="popB="popB="popB="popB="popB="popB="popB="popB="popB
       n.signal=100,window.size=0.01)
 # Plot resulting admixture signal for one individual
plotsignal(AdexPCA3,ind="AD00001",popA=AdexPCA2$popA,popB=AdexPCA2$popB)
# EXAMPLE 4
 # Generate the admixture signal in terms of genetic distance
 # As in EXAMPLE 2 but with genmap specified so that signals are formulated using genetic distances
AdexPCA4 <- signal(admix\$data,popA="popA",popB="popB",populations=admix\$populations,tol=0.001,admix\$data,popA="popA",popB="popB",populations=admix\$populations,tol=0.001,admix\$data,popA="popA",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB="popB",popB",popB="popB",popB",popB",popB="popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB",popB
   n.signal=1000,window.size=0.01,genmap=admix$map[,2])
 # Plot resulting admixture signal for one individual
 plotsignal(AdexPCA4,ind="AD00001",popA=AdexPCA4$popA,popB=AdexPCA4$popB)
```

## **Description**

Produces wavelet summaries for each individual and group. Returns the wavelet variance and average block size metric (ABS).

#### Usage

### **Arguments**

x	object of class adsig.
populations	list containing a vector of individual IDs for each population in the analysis.
рорА	name of ancestral population 1 (used for forming the axes of variation). Must match one of the names in populations.
рорВ	name of ancestral population 2 (used for forming the axes of variation). Must match one of the names in populations.
ml	number of wavelet scales in the decomposition. Must not exceed $log_2(T)$ , where $T$ is the length of the signal.
type	name of the wavelet to use in the decomposition. The default, "la8", is Daubechies Least Asymmetric wavelet of length 8. Other options include "haar".
t.factor	multiplicative factor for thresholding. See paper for details. Default is 1.
fullWT	if TRUE, save the full wavelet periodogram, as well as the wavelet variance. Object size will be large. Default is FALSE.

#### **Details**

Produces wavelet summaries for objects of class adsig. The function computes the wavelet variance for each individual and population, extracts the informative wavelet variance based on levels observed in the ancestral populations, and computes summary measures of average block size metric (ABS) and peak wavelet scale for each individual and population.

See waveslim documentation for details of the modwt function and alternative wavelet options.

#### Value

The code returns a list with the following components:

n.ind	number of individuals in the analysis.
n.group	number of groups in the analysis.
rv.ind	matrix of dimension $nxml$ , returning the raw wavelet variance for each individual.
rv.group	matrix of dimension $n.groupxml$ , returning the raw wavelet variance for each group.
threshold	vector of length $ml$ , returning threshold values for each wavelet scale.

iv.ind	matrix of dimension $nxml$ , returning the informative (thresholded) wavelet variance for each individual.
iv.group	matrix of dimension $n.groupxml$ , returning the informative (thresholded) wavelet variance for each group.
abs.ind	vector of length $n$ , returning the average block size metric (ABS) for each individual.
abs.group	vector of length n.group, returning the average block size metric (ABS) for each group.
pws.ind	vector of length $n$ , returning the peak wavelet scale for each individual.
pws.group	vector of length n. group, returning the peak wavelet scale for each group.
wtmatrix	$(onlyif{\rm fullWT}={\rm TRUE}).ArrayofdimensionTxnxml, containingsquaredwaveletcoefficientsforeachindividual.$
wtmatrix.group	(only if fullWT = TRUE). Array of dimension $Txn.groupxml$ , squared wavelet coefficients, averaged for each group.

## Author(s)

Jean Sanderson

#### References

Sanderson J, H Sudoyo, TM Karafet, MF Hammer and MP Cox. 2015. Reconstructing past admixture processes from local genomic ancestry using wavelet transformation. *Genetics* 200:469-481. https://doi.org/10.1534/genetics.115.176842

#### See Also

signal

## **Examples**

```
data(admix)
# Generate the admixture signal
AdexPCA <- signal(admix$data,popA="popA",popB="popB",populations=admix$populations,
    tol=0.001, n.signal=NULL)

# Compute wavelet summaries
WSN <- wavesum(AdexPCA,populations=admix$populations,popA="popA",popB="popB")

# Plot raw wavelet variance for each population
barplot(WSN$rv.group[3,],ylim=c(0,0.9),col="red", names.arg=1:11,border=NA)
barplot(WSN$rv.group[1,],ylim=c(0,0.9),col="green3",names.arg=1:11,border=NA,add=TRUE)
barplot(WSN$rv.group[2,],ylim=c(0,0.9),col="blue", names.arg=1:11,border=NA,add=TRUE)
legend("topright",c("popA","popB","popAB"),col=c(3,4,2),pch=15)
box()

# Plot informative wavelet variance for admixed population
barplot(WSN$iv.group[3,],ylim=c(0,0.15),col="red",names.arg=1:11,border=NA)</pre>
```

```
ABS <- round(WSN$abs.group[3],2) text(11,0.13,paste("ABS=",ABS)) box()
```

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